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Note

A new cucurbitacin from *Bolbostemma paniculatum* Franguent

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A new cucurbitacin with an unusual ring A, isocucurbitacin D 25-*O*-acetate (1), was isolated from *Bolbostemma paniculatum* Franguent together with one known compound, cucurbitacin E (2). The structure of new compound was established by spectroscopic methods.

Keywords: *Bolbostemma paniculatum* (Maxim.) Franguent; Cucurbitacin; Isocucurbitacin

1. Introduction

Bolbostemma paniculatum (Maxim.) Franguent is one of the Chinese folk medicines often used for the treatment of tumours as well as for detoxification. It is distributed in most provinces of China. Many triterpenoids have been isolated from related species of the same family over the past decade. Cucurbitacins are a special group of triterpenoids having a cucurbitane skeleton characterised by a 19 (10 → 9β) abeo-10α-lanost-5-ene. Most of the cucurbitacins are tetracyclic, but some representatives have an extra ring due to formal cyclisation between C-16 and C-24 (cucurbitacins S and T) [1,2]. Certain cucurbitacins have been discovered in the form of glycosides and some of them lack C-11 carbonyl function [3]. Biologically, they exhibit a wide range of activities including cytotoxicity and anti-tumour effects [4–6]. Chemically, cucurbitacins are classified according to the functionalities in rings A and C, side chain modifications, as well as stereochemical considerations. In this paper we report the isolation and structural elucidation of these two cucurbitacins.

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2. Results and discussion

Chromatographic fractionation of the chloroform extract of *Bolbostemma paniculatum* Frangent on silica gel column followed by HPLC separation yielded two cucurbitacins, **1** and **2**, which showed brown colour with H₂SO₄ spray reagent.

Compound **1** was obtained as a white powder, mp 228–230°C, $[\alpha]_D^{20} + 62.5$ (*c* 0.52, CHCl₃). Its molecular formula was determined as C₃₂H₄₆O₈ by HRESI-MS *m/z* 581.3090 [M + Na]⁺. The IR spectrum indicated the presence of hydroxyls, carbonyls, and olefinic functionalities. The ¹H NMR (600 MHz, CDCl₃) spectrum showed a sharp singlet at δ 3.91 assigned to the α-hydroxylated methine group, indicating the presence of the isocucurbitacin skeleton with 2-keto-3-α hydroxy system in ring A [7]. Three olefinic signals at δ 5.94 (d, *J* = 5.28 Hz, H-6), 6.43 (d, *J* = 15.60 Hz, H-23) and 7.05 (d, *J* = 15.60 Hz, H-24) and an AB system at δ 2.64 (H-12β, d, *J* = 14.64 Hz) and 3.12 (H-12α, d, *J* = 14.64 Hz) were observed in the ¹H NMR spectrum. The signal at δ 4.34 was assigned to H-16β [8]. In addition, **1** showed nine methyl singlets at δ 0.81–2.00

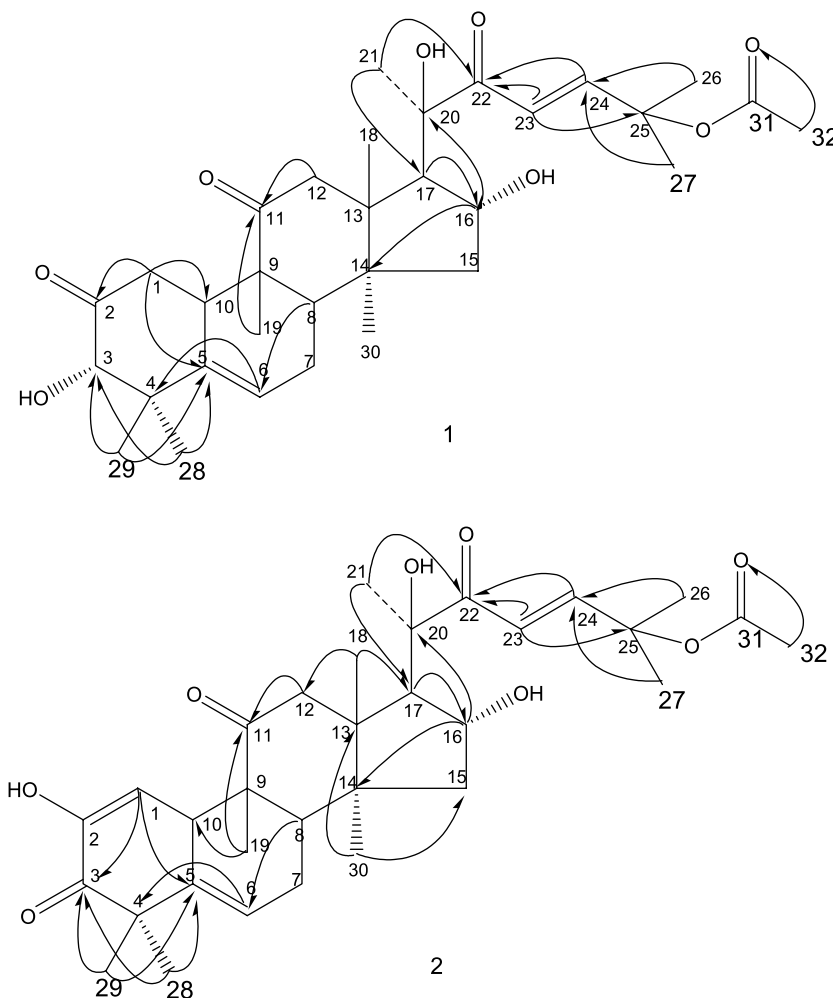


Figure 1. Key HMBC correlations of compounds **1** and **2**.

among which one methyl singlet at δ 2.00 was assignable to the 25-*O*-acetate group (C-31) [8,9]. ^{13}C NMR experiments showed 32 carbon signals including four carbonyls, nine methyls and four olefinic carbons. Nine methyls were located by means of HMQC and HMBC experiments (figure 1). In the HMBC spectrum two protons at δ 2.41 (H-1 β) and 2.23 (H-1 α) showed long-range correlations with the carbonyl carbon at δ 210.6 (C-2). The complete structural elucidation of **1** was derived from the chemical shifts and coupling constant of the ^1H NMR and ^{13}C NMR spectra, and detailed spectral analysis of HMQC, HMBC experiments (table 1) and comparison with the data of the literature [10–12]. Therefore **1** was characterised as 19-norlanosta-5,23-diene-2,11,22-trione-25-(acetyloxy)-3 α ,16 α ,20 β -trihydroxy-9-methyl.

Compound **2** was one known compound identified by spectroscopic methods and comparison with the data of the literature [13,14].

3. Experimental

3.1 General experimental procedures

The NMR data were recorded on a Bruker AV-600 spectrometer (600 MHz for ^1H and 150 MHz for ^{13}C) in CD_3Cl with TMS as internal standard. The IR spectra were recorded on

Table 1. ^1H NMR (600 MHz) and ^{13}C NMR (150 MHz) data of compound **1**.

<i>H</i>	δ_H	<i>C</i>	δ_C
H-1 α	2.23 (m)	1	38.8
H-1 β	2.41 (m)	2	210.6
H-3	3.91 (s)	3	80.2
H-6	5.94 (d, $J = 5.28$)	4	46.7
H-7 α	1.99 (m)	5	138.1
H-7 β	2.50 (m)	6	121.9
H-8	2.04 (d, $J = 7.68$)	7	23.8
H-10	2.74 (br d, $J = 12.06$)	8	42.7
H-12 α	3.12 (d, $J = 14.64$)	9	48.3
H-12 β	2.64 (d, $J = 14.64$)	10	36.3
H-15 α	1.86 (dd, $J = 9.36, 13.14$)	11	211.8
H-15 β	1.44 (m)	12	48.6
H-16	4.34 (m)	13	50.6
H-17	2.45 (d, $J = 7.14$)	14	47.9
H-23	6.43 (d, $J = 15.60$)	15	45.3
H-24	7.05 (d, $J = 15.60$)	16	71.3
Me-18	0.97	17	58.1
Me-19	1.18	18	19.9
Me-21	1.42	19	20.0
Me-26	1.56	20	78.1
Me-27	1.53	21	23.6
Me-28	0.81	22	202.4
Me-29	1.33	23	120.2
Me-30	1.26	24	152.1
Me-32	2.00	25	79.3
		26	25.9
		27	26.4
		28	20.9
		29	24.0
		30	18.8
		31	170.2
		32	21.9

a Bruker IFS-55 instrument (KBr). HRESI-MS spectra were measured on a Apex II FTICR mass spectrometer. Chromatography was performed with silica gel (200–300 mesh), Sephadex LH-20 and HPLC.

3.2 Plant material

The underground parts of *Bolbostemma paniculatum* (Maxim.) were offered by Changchun Huakang Pharmaceutical Company Ltd. and were identified by Professor Qi-shi Sun (Shenyang Pharmaceutical University). A sample has been deposited in the School of Chinese Medicine, Shenyang Pharmaceutical University.

3.3 Extraction and isolation

Dried and powdered underground parts of *B. paniculatum* (Maxim.) were extracted three times with EtOH. After evaporation of EtOH *in vacuo*, the concentrated extract was suspended in water and partitioned with CHCl₃, AcOEt, and n-BuOH, respectively. The CHCl₃ portion (103 g) was chromatographed on a silica gel (1030 g, Qingdao Haiyang Chemical Co. Ltd.) column and eluted with petroleum ether/acetone (100:20) to give fractions monitored by TLC. Fraction 1 (0.5 g) was rechromatographed by HPLC (MeOH/H₂O 60:40 v/v; flow rate 1 ml/min; UV detection 210 nm), to afford compounds **1** (20 mg) and **2** (16 mg).

3.3.1 Compound 1. White powder, mp 228–230°C, $[\alpha]_D^{20} + 62.5$ (*c* 0.52, CHCl₃), positive Libermann reaction; TLC: *R*_f 0.52, HPLC: *t*_R 34.5 min. HRESI-MS *m/z* 581.3090 [M + Na]⁺ (calcd for C₃₂H₄₆O₈, 581.3097); IR (KBr) cm⁻¹: 3378, 1720, 1683, 1658, 1633, 1362, 1260, 1121, 1038, 990. ¹H NMR, ¹³C NMR data: see table 1; HMBC data: see figure 1.

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